Protective effect of dexpanthenol on gentamicin-induced nephrotoxicity in rats

Neslihan Pinar¹, Oguzhan Ozcan², Esin Dogan³, Gokhan Cakirca⁴

¹ Mustafa Kemal University, Faculty of Medicine, Department of Medical Pharmacology, Hatay, Turkey
² Mustafa Kemal University, Faculty of Medicine, Department of Biochemistry, Hatay, Turkey
³ Mustafa Kemal University, Faculty of Medicine, Department of Biochemistry, Hatay, Turkey
⁴ Sanliurfa Mehmet Akif Inan Training and Research Hospital, Department of Biochemistry, Sanliurfa, Turkey

Abstract

Aim: We evaluated the protective effects of dexpanthenol (Dxp) in rats with gentamicin (Genta)-induced nephrotoxicity by assessing a panel of biochemical and histopathologic parameters.

Material Methods: Forty rats were divided randomly into the following four groups: Control group, physiological saline solution (0.5 cc intraperitoneally (i.p.) for 8 days; Dxp group, Dxp (500 mg/kg i.p.) for 8 days; Genta group, Genta (100 mg/kg, i.p.) for 8 days; and Genta+Dxp group, Genta and Dxp (100 and 500 mg/kg i.p., respectively) for 8 days.

Results: In the Genta group, the urea, creatinine, tumor necrosis factor-alpha (TNF-α), total oxidant status (TOS), oxidative stress index (OSI) and malondialdehyde (MDA) levels were significantly higher and the catalase (CAT) and glutathione peroxidase (GSH-Px) activities were significantly lower than those in the control group. In the Genta+Dxp group, the urea, creatinine, and TNF-α, TOS, OSI and MDA levels were significantly lower and the CAT and GSH-Px activities were significantly higher than those in the Genta group. Histopathological investigation showed severe tubular necrosis in the Genta group, which was of lesser severity in the Genta+Dxp group.

Conclusion: The biochemical and histopathologic results of this study indicate that Dxp can ameliorate Genta-induced nephrotoxicity.

Keywords: Oxidative stress; antioxidant; gentamicin; nephrotoxicity; dexpanthenol.

INTRODUCTION

Gentamicin (Genta) is an aminoglycoside antibiotic frequently used against infections caused by Gram negative bacteria. However, clinical use of Genta is limited by its nephrotoxicity and ototoxicity (1). High doses of Genta cause nephrotoxicity due to its accumulation in epithelial cells of the proximal renal tubules (2). Genta in rats leads to histopathological alterations such as edema, basal membrane interruption, apoptosis, and tubular necrosis (3). The mechanism of Genta-induced nephrotoxicity is unclear but involves a number of factors including reactive oxygen species (ROS), inflammation, and apoptosis (4,5). Increased production of ROS, depressed antioxidant defenses, and excessive levels of proinflammatory cytokines play a key role in the pathogenesis of Genta-induced nephrotoxicity (6,7). Thus, various antioxidants and anti-inflammatory agents have been used to treat or prevent Genta-induced nephrotoxicity (8,9).

Dexpanthenol (Dxp), an alcohol derivative of pantothenic acid (PA), is oxidized to PA in tissues; this increases glutathione, coenzyme-A, and ATP synthesis (10). Dxp plays an important role in both cellular defense against oxidative stress and the inflammatory response (11,12). Hence, we investigated the antioxidant and anti-inflammatory effects of Dxp in rats with Genta-induced nephrotoxicity by assessing a panel of biochemical and histopathological parameters.

MATERIAL and METHODS

Chemicals
Genta sulfate (Genta ampoule 80 mg, I.E. Istanbul, Turkey) and Dxp (Bepanthene ampoule 500 mg/2 mL, Bayer) were used in this study.
Animals
Forty male Wistar albino rats weighing 250–300 g were used in this study. The Mustafa Kemal University Experimental Animals Local Ethics Committee approved the study (Approval No., 2015/5-4). The experiment was carried out in Mustafa Kemal University Experimental Researches Application and Research Center, Hatay, Turkey. The rats were fed ad libitum and housed in appropriate cages, each containing five rats. The rats were exposed to a 12/12 h light/dark cycle at 20–22°C and 50–55% humidity.

Experimental Design
Forty rats were divided into four groups. Physiological saline was injected (0.5 mL intraperitoneally (i.p.) for 8 days to the rats in the control group. Dxp (500 mg/kg i.p.) was injected for 8 days to the rats in the Dxp group (13,14).The rats in the Genta group received injections of Genta (100 mg/kg i.p.) for 8 days (15).The rats in the Genta+Dxp group were injected with Genta and Dxp (100 and 500 mg/kg i.p., respectively) for 8 days.

Twenty-four hours after the last injection, the rats were anesthetized with ketamine hydrochloride (60 mg/kg i.p.) and xylazine hydrochloride (10 mg/kg i.p.), subjected to intracardiac blood sampling, and decapitated. A midline abdominal incision was made, and the left kidney was removed and transferred to the laboratory in 10% neutral formaldehyde solution in a closed labeled bottle for histopathological evaluation. The right kidney was removed, covered with labeled aluminum foil, and transferred in liquid nitrogen to the biochemistry laboratory. Blood samples were centrifuged at 1,500 rpm for 10 minutes and serum samples were stored at −80°C until required. The biochemical parameters assessed were the serum urea, creatinine, total antioxidant status (TAS), total oxidant status (TOS), and tumor necrosis factor-alpha (TNF-α) levels. The malondialdehyde (MDA) level, catalase (CAT) activity, and glutathione peroxidase (GSH Px) activity in renal tissue were also evaluated.

Biochemical Analysis
The TOS and TAS levels were determined by the method of Erel (16,17). The oxidative stress index (OSI) was defined as the TOS to TAS ratio. The serum levels of urea, creatinine, Na, and K were measured spectrophotometrically using an Architect c8000 analyzer (Abbott Diagnostics). The serum TNF-α level was measured using an enzyme-linked immunosorbent assay kit (Awareness Technology Inc. ChroMate ELISA) and expressed as pg/mL protein. CAT activity was evaluated by the method of Aebi (18). GSH-Px activity was measured according to the recommendations of Paglia and Valentine (19). The MDA level was evaluated by the method of Draper and Hadley (20).

Histopathological Analysis
Renal tissue samples were fixed in 10% neutral formaldehyde solution and embedded in paraffin. Paraffin sections (4 μm thickness) were cut using a microtome and stained with hematoxylin and eosin. The sections were visualized using a light microscope. Tubular necrosis, tubular degeneration, congestion, tubular hypertrophy, and perivascular inflammation were evaluated by light microscopy. A predetermined scoring system was used for histopathological evaluation (21). Tissue damage was scored as follows: +, slight; ++, moderate; and +++, severe.

Statistical analysis
Statistical analyses were performed using SPSS ver. 21 (SPSS Inc., Chicago, IL, USA). The values are expressed as mean ± SEM. The groups were compared by one-way analysis of variance (ANOVA) followed by Tukey’s test. A value of p < 0.05 was considered indicative of statistical significance.

RESULTS
Biochemical results
The results of the oxidative stress and biochemical parameters in rats with Genta-induced nephrotoxicity are shown in Table 1 and 2. The urea, creatinine and TNF-α levels were significantly higher in the Genta group than in the control group and were significantly lower in the Genta+Dxp group than in the Genta group (p < 0.05). The serum Na level did not differ significantly among the groups, but the serum K level was significantly lower in the Genta+Dxp group than in the Dxp group (p < 0.05) (Table 1).

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Control</th>
<th>Dxp</th>
<th>Genta</th>
<th>Genta+Dxp</th>
</tr>
</thead>
<tbody>
<tr>
<td>TNF-α (pg/mL protein)</td>
<td>115.5± 8.1</td>
<td>335.8±84.8</td>
<td>899.3±181.3*</td>
<td>386.3 ±65.4**</td>
</tr>
<tr>
<td>Urea (mg/dL)</td>
<td>20.3±0.25</td>
<td>20.7±0.37</td>
<td>130.8±29.8*</td>
<td>61.9 ± 11.9**</td>
</tr>
<tr>
<td>Creatinine (mg/dL)</td>
<td>0.5±0.01</td>
<td>0.49±0.0</td>
<td>2.55±0.6*</td>
<td>1.04 ± 0.24**</td>
</tr>
<tr>
<td>Na (mmol/L)</td>
<td>135.5±1.4</td>
<td>138.2±0.4</td>
<td>134.7±1.57</td>
<td>139.1 ± 0.89</td>
</tr>
<tr>
<td>K (mmol/L)</td>
<td>5.67±0.3</td>
<td>5.72±0.17</td>
<td>5.32±0.27</td>
<td>4.6 ± 0.21***</td>
</tr>
</tbody>
</table>

Data were expressed as Mean ± SEM. TNF-α: Tumor necrosis factor-alpha, Na: Sodium, K: Potassium.
*p<0.05 vs. Control group
**p<0.05 vs. Genta group
***p<0.05 vs. Dexa group
The TOS, OSI and MDA levels were higher in the Genta group than in the control group and were significantly lower in the Genta+Dxp group than in the Genta group (p < 0.05). However, the TAS did not differ significantly among the four groups (p > 0.05). The CAT and GSH-Px levels were lower in the Genta group than in the control group and were significantly higher in the Genta+Dxp group than in the Genta group (p < 0.05) (Table 2).

Histological results
The renal tissues were histopathologically evaluated for tubular necrosis, tubular degeneration, glomerular atrophy and hypertrophy, and perivascular inflammation. The control and Dxp groups had normal renal tissue (Figure 1A and 1B). The rats in the Genta group had marked proximal tubular necrosis, tubular degeneration, perivascular inflammation (interstitial nephritis and pyelonephritis), and marked glomerular hypertrophy (Figure 1C). The rats in the Genta+Dxp group showed mild tubular necrosis, glomerular atrophy, and perivascular inflammation and moderate tubular degeneration and glomerular hypertrophy (Figure 1D).

Table 1. Effects of dexpanthenol (Dxp) on biochemical parameters

<table>
<thead>
<tr>
<th>Parameters</th>
<th>Control</th>
<th>Dxp</th>
<th>Genta</th>
<th>Genta+Dxp</th>
</tr>
</thead>
<tbody>
<tr>
<td>TAS (µmol Trolox Eq/g protein)</td>
<td>0.92 ± 0.07</td>
<td>0.94 ± 0.03</td>
<td>0.86 ± 0.07</td>
<td>0.98 ± 0.03</td>
</tr>
<tr>
<td>TOS (µmol H2O2 Eq/g protein)</td>
<td>12.6 ± 1.00</td>
<td>16.9 ± 2.00</td>
<td>31.5 ± 4.84*</td>
<td>19.4 ± 2.28**</td>
</tr>
<tr>
<td>OSI (H2O2/Trolox)</td>
<td>1.39 ± 0.14</td>
<td>1.92 ± 0.24</td>
<td>3.65 ± 0.49*</td>
<td>1.98 ± 0.22**</td>
</tr>
<tr>
<td>MDA</td>
<td>0.46 ± 0.02</td>
<td>0.45 ± 0.02</td>
<td>1.05 ± 0.04*</td>
<td>0.71 ± 0.03**</td>
</tr>
<tr>
<td>CAT</td>
<td>0.07 ± 0.00</td>
<td>0.07 ± 0.02</td>
<td>0.03 ± 0.00*</td>
<td>0.04 ± 0.00**</td>
</tr>
<tr>
<td>GSH-Px</td>
<td>0.32 ± 0.01</td>
<td>0.30 ± 0.01</td>
<td>0.18 ± 0.00*</td>
<td>0.23 ± 0.01**</td>
</tr>
</tbody>
</table>

Data were expressed as Mean ± SEM. TAS: Total antioxidant status, TOS: Total oxidant status, OSI: Oxidative stress index, MDA: Malondialdehyde, CAT: Catalase, GSH-Px: glutathione peroxidase.

*p<0.05 vs. Control group
**p<0.05 vs. Genta group

DISCUSSION
We evaluated the protective effect of Dxp on Genta induced nephrotoxicity. The results showed that Dxp ameliorated the renal injury caused by high-dose Genta by decreasing oxidative stress and the inflammation.

Clinical use of aminoglycosides is restricted by their nephrotoxicity, which is caused by generation of ROS (5). In renal pathology, NADPH oxidases, together with the mitochondrial respiratory chain, are the major sources of ROS such as the superoxide anion (O2⁻), hydrogen peroxide (H₂O₂), and the hydroxyl radical (•OH) (22). Abnormal production of ROS leads to cellular injury and necrosis by inducing membrane lipid peroxidation, protein denaturation, and damage to DNA. The changes in renal function caused by lipid peroxidation are considered to be the initial event in the Genta-induced nephrotoxicity injury cascade (5,23). Genta increases ROS production, reduces the antioxidant capacity, and increases lipid peroxidation (24,25). Genta-induced renal injury can be prevented by various agents (6,9). In this study, we investigated for the first time the therapeutic effects of Dxp, which has antioxidant and anti-inflammatory activity, on Genta-induced nephrotoxicity.

Genta causes nephrotoxicity by decreasing the glomerular filtration rate and increasing the serum urea and creatinine levels (26,27). In our study, Genta led to a significant...
increase in the serum urea and creatinine levels, whereas Genta+Dxp induced a significant decrease in the serum urea and creatinine levels. These results suggest a role for Dxp in preventing Genta-induced nephrotoxicity.

Inflammatory reactions, which are characterized by leukocyte infiltration into the injury site, increase the production of proinflammatory cytokines, such as TNF-α, and activation of nuclear factor-kB, also play an important role in the nephrotoxicity of Genta (5,28). In our study, the TNF-α level was significantly higher in the Genta group and significantly lower in the Genta+Dxp group compared to the Genta group. Thus, the anti-inflammatory activity of Dxp suppressed renal inflammation.

Excess ROS production caused by the nephrotoxic effect of Genta damages cells by interacting with membrane lipids. MDA is a product of lipid peroxidation by ROS (22). In our study, as in prior reports, Genta led to a significant increase in the MDA level (29,30). In contrast, the MDA level was significantly lower in the Genta+Dxp group compared to the Genta group.

GSH-Px and CAT are enzymatic antioxidants that protect cells against the negative effects of free radicals, for example, by converting H2O2 into H2O (31). In this study, the GSH-Px and CAT activities were lower in the Genta group than in the control group, as reported previously (23,26). On the other hand, GSH-Px and CAT activities were higher in the Genta+Dxp group than in the Genta group.

We also measured the TAS, TOS, and OSI to evaluate the oxidant and antioxidant status in Genta-induced nephrotoxicity. The TOS and the OSI value were significantly higher in the Genta group compared to the control group; this increase was reversed by simultaneous administration of Dxp with Genta. These results indicate that Dxp reduces the level of oxidative stress induced by Genta. Collectively, our results showed that Dxp administration increases the activities of GSH-Px and CAT and decreases the TOS and the OSI value and the level of MDA. This effect is likely to be mediated by the antioxidant activity of Dxp, which protects the kidney against free oxygen radicals. Similarly, Dxp protected against ischemia-reperfusion-induced renal injury in rats (32,33). Thus, Dxp has antioxidant activity and reduces the generation of free radicals.

Genta-induced nephrotoxicity is morphologically characterized by proximal tubule epithelial desquamation, epithelial edema, glomerular congestion, necrosis, and tubular casts (34). In this study, the renal tissue of the rats in the Genta group exhibited marked glomerular hypertrophy, perivascular inflammation, congestion, tubular degeneration, and tubular necrosis. Thus, Genta induced severe renal injury. In the Genta+Dxp group, however, Genta induced moderate renal injury, suggesting that Dxp ameliorated the tubular injury caused by Genta. Also, the histopathological changes were consistent with the biochemical findings.

CONCLUSION

Our results showed that Genta-induced nephrotoxicity in rats was ameliorated by the anti-inflammatory and antioxidant effects of Dxp. We believe that Dxp can be used safely in clinical practice to prevent Genta-induced kidney damage. However, further studies of the protective effect of Dxp on Genta-induced nephrotoxicity are required.

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