The investigation of lipoprotein-related phospholipase A2 (LP-PLA2) V279F mutation in coronary artery disease

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Abstract

Aim: Despite advances in the treatment of cardiovascular diseases, coronary artery disease (CAD) is the leading cause of death worldwide. The media layer of normal and diseased arteries contains lipoprotein-associated phospholipase A2 (LP-PLA2), which is mostly made by mast cells, T lymphocytes, and monocytes. Our research aimed to define how the LP-PLA2 and V279F gene polymorphisms are related to CAD.

Materials and Methods: This study included 109 CAD patients and 71 controls. Fast- ing blood glucose (FBG) and lipid profiles were measured via the enzymatic colorimetric technique. LP-PLA2 levels were detected with ELISA. The genomic DNAs were isolated from whole blood. Real-time polymerase chain reaction (RT-PCR) with the V279F mutation detection kit was used to identify the LP-PLA2 V279F mutation.

Results: In the CAD group compared to controls, serum LP-PLA2 levels were higher (p < 0.001). While FF (mutant) genotypes and VF (heterozygous) genotypes were absent in both groups, the LP-PLA2 V279F mutation was discovered as the VV (wild) genotype in both the CAD group and controls.

Conclusion: As a result, we determined that the LP-PLA2 V279F mutation cannot be considered a genetic risk factor for CAD, but that LP-PLA2 elevation may be an important parameter for CAD. Keywords: LP-PLA2, Coronary artery disease, V279F.

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tion and both have pro-apoptotic effects on macrophages and also have high solubility, so they can easily diffuse into atheroma and act on various cell types, which have a role in atherosclerosis. LPC is a potent chemoattractant for monocytes and T cells, inducing apoptosis in smooth muscle cells promoting endothelial dysfunction, and stimulating macrophage proliferation. Foam cells and activated macrophages produce more Lp-PLA2 [9-13]. A higher oxidation degree of LDL indicates higher LPC concentration. Studies have displayed that oxidized LDL increases the risk of atherosclerosis. In addition, OxFFA and LPC stimulate the formation of adhesion particles and cytokines by plaque-based macrophages, endothelial cells, and leukocytes [14,15].

The Lp-PLA2 gene is based at the 6q21.2-p12 region of the chromosome and consists of 12 exons. The V279F mutation, first identified in Lp-PLA2, is a single point mutation in exon 9 at nucleotide 994, with Thymine (G994T) replaced by Guanine. This nucleotide change causes the formation of Valine 279 Phenylalanine (V279F) in the mature protein and a lack of activity in the Lp-PLA2 enzyme. It has been defined that the molecular basis of Lp-PLA2 enzyme deficiency is caused by a single point mutation of V279F close to the active site of the enzyme [13,16].

The exposure of patients without traditional cardiovascular risk factors to cardiovascular events has prompted researchers to seek new markers that can better predict cardiovascular risks. Therefore, we examined Lp-PLA2 levels and Lp-PLA2 V279F single point mutation in relation to CAD to find whether Lp-PLA2 is a new risk factor.

Materials and Methods

Study subjects

Our study group consisted of 180 individuals, 46 females and 134 males, presented to the Mersin University Hospital Cardiology outpatient clinic with chest pain and underwent coronary angiography between September 2010 and September 2011. According to the angiography results, 109 subjects with a minimum of one coronary artery stenosis over 70% formed the patient group, and 71 subjects without coronary artery lesions formed the control group. Patients with a history of coronary artery bypass graft surgery, inflammatory disease, infectious disease, hematological disease, autoimmune disease, kidney disease, liver disease, or malignancy, were excluded from the study. Power analysis was performed using G Power 3.0.10 for Windows. In order for the difference between the patient and control groups in terms of the Lp-PLA2 enzyme activity to be found statistically significant, at least 100 cases in each group are sufficient with a Type I error of 0.05 and 80% power. Fifteen subjects who did not meet the inclusion criteria in the control group were excluded from the study, while 13 subjects in the control group refused to participate in the study.

The Mersin University University Clinical Research Ethics Committee approved the study (05.15.2009, 2009/105) and informed consent was acquired from all individuals enrolled in the research.

Blood sampling

During the angiography procedure, 10 mL of blood was taken from the femoral artery, and it was divided into tubes containing ethylenediaminetetraacetic acid (EDTA) for DNA extraction and biochemistry tubes for separating the sera. Whole blood taken to detect the Lp-PLA2 V279F mutation in the patient and control groups was stored at +4 °C for DNA isolation until analysis. Peripheral venous blood taken into biochemistry tubes for lipid profile and fasting blood glucose (FBG) levels were centrifuged at 4000 rpm for 10 minutes and serum was allocated and studied in a Cobas 501 autoanalyzer (Roche Diagnostics, Mannheim GmbH, Germany). Serum Lp-PLA2 levels were measured by the enzyme-linked immunosorbent assay (ELISA) method using the ELISA (Cat. No. E90867Hu, USCN Life Science Inc., China) kit.

DNA extraction and genotyping

To perform genetic analysis, peripheral blood was drawn into tubes containing EDTA. DNA extraction was performed from circulating leukocytes by utilizing a high pure polymerase chain reaction (PCR) template preparation kit (Roche Diagnostics, GmbH, Mannheim, Germany, catalog no: 1 796 828). DNA was extracted according to the methodology laid out by Roche Diagnostics GmbH. For the DNA extraction step of this procedure, we employed proteinase K, binding buffer, inhibitor removal buffer, isopropanol, wash buffer, and elution buffer. Lp-PLA2 V279F gene polymorphism was analyzed by utilizing LightCycler mutation detection kits (TIB MOLBIOL) in real-time-PCR with a LightCycler instrument (Roche Diagnostics, Mannheim, Germany). The primer sequences were:

- forward primer: 5’_CAAATA ggACTCTTATATg_3’;
- reverse primer: 5’_gCTTTACTATCTTATTTTCTTACCTg_3’
- and the probes sequences were detection probe: 5’_AgTCTgAATAACCgTTgCTCC--FL_3’
- and anchor probe: 5’_LC-CCAAAA gAAtgTCCAATTACTgCTATTTTT--PH_3’.

Statistical analysis

The obtained data were analyzed using IBM Statistical Package for the Social Sciences Demo Version 20.0 package (IBM SPSS Inc. Free Download, Chicago, Illinois, USA) program. The Kolmogorov-Smirnov and Shapiro-Wilk tests were used to evaluate the normal distribution, and the Levene test was used to examine the homogeneity of variances. Quantitative data conforming to the normal distribution were presented as mean ± standard deviation and compared with independent samples t-test or One-Way ANOVA depending on the number of groups compared. Qualitative data were summarized with frequency and percentage, and the chi-square test was used to analyze qualitative data. The logistic regression analysis was performed to determine the relationship between categorical variables. A p-value of <0.05 was considered a statistical significance level.
Results
The study included 109 healthy individuals with a mean age of 62.6±9.4 years (17 females and 92 males) and 71 individuals with CAD with a mean age of 55.8±11.6 years (29 females and 42 males). The distribution and descriptive information of risk factors in CAD and control groups are given in Table 1. The age difference between groups was statistically significant (p=0.001). Males were 3.7 times more likely to have CAD than females and represented a higher proportion of CAD patients compared to controls (OR: 3.73, 95% CI: 1.85-7.53; p=0.001). Smokers had a 2-fold (p=0.023) risk of CAD and those with a family history of CAD had a 2.89-fold (p=0.002) risk of CAD. The presence of hypertension (p=0.092) and diabetes (p=0.16), which are considered among the risk factors for CAD, was not a risk factor for CAD.

Compared to controls, serum FBG levels were higher in the CAD group (p=0.01) and due to the usage of statin group medications with cholesterol-lowering effects in the CAD group, there was no significant difference between the two groups’ cholesterol levels, which is the main risk factor for CAD (p=0.27) (Table 2). Lp-PLA2 levels of both groups are presented in Figure 1. The Lp-PLA2 level was 90.6±87 ng/mL in the control group, while it was 140±119 ng/mL in the CAD group. Serum Lp-PLA2 levels were higher in the CAD group than in controls (p=0.0029). Lp-PLA2 levels were 102.31±94.74, 142.44±123.9, and 173.17±122.3 ng/mL in vessel 1, vessel 2, and vessel 3, respectively (Figure 2). Levels of Lp-PLA2 increased according to the number of occluded coronary arteries, and patients with 3 involved vessels and above had the highest Lp-PLA2 levels. A statistically significant difference was detected only between those with 3 involved vessels and above and those with 1 involved vessel (p=0.016). In both groups, only the Lp-PLA2 V279V (wild) genotype was detected as 100%. Lp-PLA2 V279F heterozygous and homozygous mutant genotypes were not detected.

Discussion
Platelet-activating factor acetylhydrolase (PAFAH), also known as LP-PLA2, has a major role in the pathogenesis of atherosclerosis. In our findings, the Lp-PLA2 level was significantly higher in the CAD (p=0.002). When the Lp-PLA2 levels were compared according to the number of occluded coronary arteries, we found that the number of CAD patients with 3 or more occluded coronary arteries was higher than those with one occluded coronary artery (p=0.016). In the literature, we found no study comparing the number of occluded coronary arteries and Lp-PLA2 levels. Many researchers have suggested that Lp-PLA2 may be a new inflammatory marker as an independent risk factor associated with CAD [17,18]. Koenig et al. [19] found a high level of Lp-PLA2 in CAD, independent of other cardiac risk markers. Kicchel et al. [20] found levels of Lp-PLA2 one standard deviation (SD) higher in the CAD group compared to controls.

In a prospective study evaluating atherosclerosis in more than 12 000 healthy middle-aged individuals over 6 years, Ballantyne et al. [21] reported that CRP and levels of Lp-PLA2 were significantly related to CAD. Caslake et al. [22] found higher serum Lp-PLA2 in 94 CAD patients compared to 54 controls. Wang et al. [23] reported that patients with severe stenosis in intracranial atherosclerosis had higher Lp-PLA2 levels. Xi et al. [24] detected elevated levels of Lp-PLA2 related to the risk of cardiovascular events and death in a middle-aged population. Lp-PLA2 levels can be affected by various factors related to the severity, phase (acute vs. chronic), and process of the disease [17, 19-25]. In order to identify whether the change in Lp-PLA2 activity is the result of an underlying cause of atherosclerotic disease or potentially a reaction to atherosclerotic changes, it is also critical to identify the patient’s Lp-PLA2 genotype. The Lp-PLA2 V279F genotype...
Table 1. Distribution of risk factors and descriptive information in control and CAD groups.

<table>
<thead>
<tr>
<th>Age</th>
<th>Control Group n=71</th>
<th>CAD Group n=109</th>
<th>Odd’s Ratio</th>
<th>Confidence Interval (%95)</th>
<th>p-value</th>
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<tbody>
<tr>
<td>Gender</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Female</td>
<td>29 (40.8)</td>
<td>17 (15.6)</td>
<td>-----</td>
<td>3.73</td>
<td>1.85-7.53</td>
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<tr>
<td>Male</td>
<td>42 (59.2)</td>
<td>92 (84.4)</td>
<td>0.001</td>
<td></td>
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<tr>
<td>Diabetes (Type 2)</td>
<td></td>
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<td></td>
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<tr>
<td>-</td>
<td>55 (77.5)</td>
<td>74 (68.0)</td>
<td>-----</td>
<td>0.16</td>
<td></td>
</tr>
<tr>
<td>+</td>
<td>16 (22.5)</td>
<td>35 (32.0)</td>
<td></td>
<td></td>
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<tr>
<td>Hypertension</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>-</td>
<td>33 (46.5)</td>
<td>37 (32.1)</td>
<td>-----</td>
<td>0.092</td>
<td></td>
</tr>
<tr>
<td>+</td>
<td>38 (53.5)</td>
<td>72 (67.9)</td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Smoking</td>
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<td>-</td>
<td>54 (76.1)</td>
<td>67 (61.5)</td>
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<td>1.02-3.88</td>
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<tr>
<td>+</td>
<td>17 (23.9)</td>
<td>42 (38.5)</td>
<td>2.00</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Family History</td>
<td></td>
<td></td>
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<tr>
<td>-</td>
<td>54 (72.6)</td>
<td>57 (52.3)</td>
<td>-----</td>
<td>0.002</td>
<td>0.473-1.86</td>
</tr>
<tr>
<td>+</td>
<td>17 (27.4)</td>
<td>52 (47.7)</td>
<td>2.89</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Number of occluded</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0</td>
<td>71</td>
<td>----</td>
<td>----</td>
<td></td>
<td>0.106</td>
</tr>
<tr>
<td>1</td>
<td>----</td>
<td>31 (30.0)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>----</td>
<td>49 (31.0)</td>
<td></td>
<td></td>
<td>0.291</td>
</tr>
<tr>
<td>3 and above</td>
<td>----</td>
<td>29 (39.0)</td>
<td></td>
<td></td>
<td>0.016</td>
</tr>
</tbody>
</table>

Continuous variables are given as mean±standard deviation. n: Number of samples, p: Significance between group. CAD: Coronary artery disease.

Table 2. Fasting Blood Sugar and lipid profile levels of control and CAD groups.

<table>
<thead>
<tr>
<th></th>
<th>Control Group</th>
<th>CAD Group</th>
<th>p-value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Fasting Blood Sugar*</td>
<td>99.87±19.99</td>
<td>112.89±45.44</td>
<td>0.01</td>
</tr>
<tr>
<td>Total Cholesterol*</td>
<td>183.07±50.88</td>
<td>175.52±39.87</td>
<td>0.27</td>
</tr>
<tr>
<td>HDL-Cholesterol*</td>
<td>42.05±12.02</td>
<td>40.64±10.07</td>
<td>0.41</td>
</tr>
<tr>
<td>LDL-Cholesterol*</td>
<td>99.11±35.44</td>
<td>111.6±41.59</td>
<td>0.56</td>
</tr>
<tr>
<td>VLDL-Cholesterol*</td>
<td>31.18±18.91</td>
<td>31.51±19.45</td>
<td>0.91</td>
</tr>
<tr>
<td>Triglyceride*</td>
<td>155.91±94.59</td>
<td>170.00±88.88</td>
<td>0.15</td>
</tr>
</tbody>
</table>

* Continuous variables are given as mean±standard deviation, and concentration units are mg/dl. p: Significance between groups.

distribution differs between races. In the study in which more than 2000 individuals participated in South America and Europe, no heterozygous or homozygous genotype was detected [26]. The V279F mutation has primarily been found in the Japanese population. Although it was later detected in ethnic groups such as Turks and Kyrgyz, it was not detected in Caucasians. The V279F mutation is associated with increased Lp-PLA2 levels in 27% of heterozygous and 4% of homozygous Japanese individuals due to a defect in enzyme secretion. Researchers have discovered that the V279F mutation is linked to cerebral hemorrhage, non-familial dilated hypertrophic cardiomyopathy, abdominal aortic aneurysm, atherosclerotic occlusive diseases, and stroke in men in the Japanese population. [24-28].

According to Yamada et al. [29] there was a larger prevalence of the V279F mutant allele in male Japanese patients with MI (homozygous, 2.1%; heterozygous, 33.0%) than in controls (homozygous, 2.2%; heterozygous, 21.0%). No significant correlation was found between the mutant genotype and CAD. Hiramoto et al. [30] found the V279F mutation in Japanese stroke patients as the heterozygous genotype as 39.2% and the homozygous genotype as 4.2%, and in the control group, the heterozygous genotype was 22.4% and the homozygous genotype was 3%. They found higher homozygous genotypes in the patient group than in controls and suggested that the V279F mutation may be a genetic risk factor in atherosclerotic diseases [30]. Balta et al. [27] verified the presence of the V279F mutation in non-Japanese populations. In their study, healthy 358 Turks, 100 Azeris, and 143 Kyrgyz were examined and heterozygous mutations were defined to be 3 (0.84%), 0, and 12 (8.4%), respectively, and no mutant genotype was detected among these individuals [27].

Contrary to other studies in Korea, Jang et al. [31] displayed that the homozygous F279F genotype reduced the risk of CAD in a study conducted with 532 men with CAD and 670 healthy individuals. In the study displayed by Şekuri et al [28] with 164 patients and 142 controls, they found the heterozygous mutation rate as 2.60% in the patient group, while all individuals in the controls were VV genotypes. They found no significant relationship between V279F heterozygous genotype and CAD. According to Ma et al. [32], there was no discernible difference in the frequency of the V279F genotype in controls and CAD patients.

Although there was no Lp-PLA2 V279F mutation in Caucasians, epidemiological studies have displayed that the risk of CAD increases with an increase in Lp-PLA2 levels [20-27,30,33]. However, in studies conducted in Asians, it was suggested that the V279F mutation, which reduces enzyme activity, increases the risk of stroke, CAD, and MI, and therefore LP-PA2 has an anti-atherogenic role in these diseases [29,30,34-36]. This difference between studies is due to genetic variation between the Caucasian and Asian...
populations and also due to both the proatherogenic and anti-atherogenic effects of the Lp-PLA2 enzyme.

In our study, all individuals in both CAD and control groups had the VV genotype. Balta et al. [27] found 3 heterozygous genotypes in 358 healthy Turkish individuals. Şeküri et al. [28] determined that there were only 3 heterozygous genotypes in 164 individuals with CAD. The mutant genotype was not detected in either study. In these studies, which included Turkish participants, the V279F heterozygous genotype rates were very low and the mutant genotype was not observed. In our findings, we could not detect the V279F heterozygous and mutant genotype. Therefore, studies in larger populations are required to confirm these findings in the Turkish population. All these studies have shown that Lp-PLA2 has been researched extensively. When the research data are evaluated in general, although it is generally believed that there may be an association between high levels of Lp-PLA2 and CAD, the opposite findings are also noteworthy.

Some restrictions must be noted regarding the current study. Our study’s primary limitation is its small participant pool. More extensive research is needed to address this. A significant correlation between Lp-PLA2 V279F mutation and CAD may have been seen if the size of our study group had been large enough.

Conclusion

In conclusion, the Lp-PLA2 level was significantly higher in the CAD group than in controls, while the Lp-PLA2 V279F mutation was not defined in either group. We think that this is because other mutations in the Lp-PLA2 gene such as Arg92His, Ile198Thr, Ile317N, and A379V affect Lp-PLA2 activity, and sequencing analysis with Lp-PLA2 protein expression is required to confirm these findings.

Finding

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Ethical approval

The Mersin University Ethics Committee approved the study. (05.15.2009, 2009/105).

References


